

Release Notes

MBT Filamentous Fungi Library 2023

Covering 225 species/entries (1021 MSPs)

Not for clinical diagnostic use

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1 Overview

1.1 Introduction

The MBT Filamentous Fungi Library 2023 is used for the identification of filamentous fungi. It consists of 1021 MSPs (MSP = **M**ain **S**pectrum **P**rojection = Reference Spectrum). The library strain composition consists of culture collection strains and strains isolated from clinical and environmental specimens provided by cooperation partners. All filamentous fungi were processed according to the Bruker Standard Operating Procedure (SOP) for filamentous fungi cultivation and sample preparation. The taxonomical classification is based on DNA sequencing whenever possible and on peer reviewed publications to assure a state-of-the-art mycological taxonomy.

1.2 What is new with the MBT Filamentous Fungi Library 2023?

The MBT Filamentous Fungi Library 2023 evolved from the MBT Filamentous Fungi Library Version 2022 which was revisited and resulted in 1021 MSPs covering 225 species/groups in total. For quality improvement some reference entries have been deleted. Where possible deleted entries have been replaced by new references of the same strain. To reflect the currently applicable nomenclature several existing references have been renamed.

2 Recommended Filamentous Fungi Cultivation and Sample Preparation Procedure

A dedicated protocol for filamentous fungi cultivation and sample preparation developed by Bruker is available. A copy of the procedure is included in the MBT HT Filamentous Fungi Module User Manual.

3 Species content of the MBT Filamentous Fungi Library

The MBT Filamentous Fungi Library 2023 covers 225 different filamentous fungi species/groups. Restrictions on species level identifications are given in the Matching Hints section.

Table 1: List of the 225 filamentous fungi species/groups in the MBT Filamentous Fungi Library 2023

	Species/Group
1	<i>Absidia caerulea</i>
2	<i>Absidia glauca</i>
3	<i>Acaulium acremonium</i>
4	<i>Acremonium cereale</i>
5	<i>Acremonium chrysogenum</i>
6	<i>Acremonium curvulum</i>
7	<i>Acremonium polychromum</i>
8	<i>Acremonium sclerotigenum</i>
9	<i>Actinomucor elegans</i>
10	<i>Alternaria alternata</i>
11	<i>Alternaria infectoria</i>
12	<i>Alternaria rosae</i>
13	<i>Apophysomyces elegans</i>
14	<i>Arthrimum arundinis</i>
15	<i>Arthrimum phaeospermum</i>
16	<i>Arthroderma borellii</i>
17	<i>Arthroderma ciferrii</i>
18	<i>Arthroderma cuniculi</i>
19	<i>Arthroderma curreyi</i>
20	<i>Arthroderma eboreum</i>
21	<i>Arthroderma flavescens</i>
22	<i>Arthroderma gertleri</i>
23	<i>Arthroderma gloriae</i>
24	<i>Arthroderma insingulare</i>
25	<i>Arthroderma lenticulare</i>
26	<i>Arthroderma multifidum</i>
27	<i>Arthroderma thuringiensis</i>
28	<i>Arthroderma uncinatum</i>
29	<i>Arthrographis kalrae</i>
30	<i>Aspergillus brasiliensis</i>
31	<i>Aspergillus calidoustus</i>
32	<i>Aspergillus carneus</i>
33	<i>Aspergillus clavatus</i>
34	<i>Aspergillus creber</i>
35	<i>Aspergillus flavus_oryzae_group</i>

	Species/Group
36	<i>Aspergillus fumigatus</i>
37	<i>Aspergillus iizukae</i>
38	<i>Aspergillus japonicus</i>
39	<i>Aspergillus lentulus</i>
40	<i>Aspergillus montevicensis</i>
41	<i>Aspergillus nidulans</i>
42	<i>Aspergillus niger</i>
43	<i>Aspergillus ochraceus</i>
44	<i>Aspergillus parasiticus</i>
45	<i>Aspergillus penicillioides</i>
46	<i>Aspergillus pseudoglaucus</i>
47	<i>Aspergillus pulvinus</i>
48	<i>Aspergillus ruber</i>
49	<i>Aspergillus rugulosus</i>
50	<i>Aspergillus sclerotiorum</i>
51	<i>Aspergillus sp[4]</i>
52	<i>Aspergillus sydowii</i>
53	<i>Aspergillus tamarii</i>
54	<i>Aspergillus terreus</i>
55	<i>Aspergillus tritici</i>
56	<i>Aspergillus unguis</i>
57	<i>Aspergillus ustus</i>
58	<i>Aspergillus versicolor</i>
59	<i>Aspergillus wentii</i>
60	<i>Aspergillus westerdijkiae</i>
61	<i>Aureobasidium melanogenum_pullulans</i>
62	<i>Beauveria bassiana</i>
63	<i>Blastomyces dermatitidis</i>
64	<i>Boeremia exigua</i>
65	<i>Botrytis aclada</i>
66	<i>Botrytis cinerea</i>
67	<i>Byssoschlamys fulva</i>
68	<i>Byssoschlamys nivea</i>
69	<i>Byssoschlamys spectabilis</i>
70	<i>Chaetomium globosum</i>



	Species/Group
71	<i>Chaetomium</i> sp
72	<i>Chrysosporium keratinophilum</i>
73	<i>Chrysosporium shanxiense</i>
74	<i>Cladosporium cladosporioides</i>
75	<i>Cladosporium halotolerans</i>
76	<i>Cladosporium herbarum</i>
77	<i>Cladosporium macrocarpum</i>
78	<i>Cladosporium sphaerospermum</i>
79	<i>Clonostachys rosea</i>
80	<i>Colletotrichum gloeosporioides</i>
81	<i>Coniochaeta hoffmannii</i>
82	<i>Coniochaeta luteorubra</i>
83	<i>Coniochaeta mutabilis</i>
84	<i>Cordyceps farinosa</i>
85	<i>Cunninghamella bertholletiae</i>
86	<i>Cunninghamella elegans</i>
87	<i>Curvularia</i> sp[7]
88	<i>Dichotomopilus dolichotrichus</i>
89	<i>Dichotomopilus funicola</i>
90	<i>Didymella glomerata</i>
91	<i>Didymella pomorum</i>
92	<i>Epicoccum nigrum</i>
93	<i>Epidermophyton floccosum</i>
94	<i>Exophiala dermatitidis</i>
95	<i>Fusarium avenaceum</i>
96	<i>Fusarium cerealis_culmorum_group</i>
97	<i>Fusarium chlamydosporum</i>
98	<i>Fusarium delphinoides</i>
99	<i>Fusarium dimerum</i>
100	<i>Fusarium equiseti</i>
101	<i>Fusarium graminearum</i>
102	<i>Fusarium incarnatum</i>
103	<i>Fusarium oxysporum</i>
104	<i>Fusarium petroliphilum</i>
105	<i>Fusarium poae</i>
106	<i>Fusarium proliferatum</i>
107	<i>Fusarium solani</i>
108	<i>Fusarium</i> sp
109	<i>Fusarium sporotrichioides</i>
110	<i>Fusarium verticillioides</i>
111	<i>Fusicolla aquaeductuum</i>

	Species/Group
112	<i>Histoplasma capsulatum</i>
113	<i>Lasiodiplodia</i> sp
114	<i>Lichtheimia corymbifera</i>
115	<i>Lichtheimia ramosa</i>
116	<i>Lomentospora prolificans</i>
117	<i>Metarhizium marquandii</i>
118	<i>Microascus gracilis</i>
119	<i>Microascus melanosporus</i>
120	<i>Microsporum audouinii_canis</i>
121	<i>Monascus ruber</i>
122	<i>Monilinia laxa</i>
123	<i>Mortierella acrotona</i>
124	<i>Mortierella angusta</i>
125	<i>Mortierella gamsii</i>
126	<i>Mortierella</i> sp
127	<i>Mucor amphibiorum</i>
128	<i>Mucor circinelloides</i>
129	<i>Mucor genevensis</i>
130	<i>Mucor hiemalis</i>
131	<i>Mucor indicus</i>
132	<i>Mucor lanceolatus</i>
133	<i>Mucor moelleri</i>
134	<i>Mucor racemosus</i>
135	<i>Mucor</i> sp
136	<i>Nannizzia aenigmatica</i>
137	<i>Nannizzia duboisii</i>
138	<i>Nannizzia fulva</i>
139	<i>Nannizzia gypsea</i>
140	<i>Nannizzia incurvata</i>
141	<i>Nannizzia persicolor</i>
142	<i>Nannizzia praecox</i>
143	<i>Neoscytalidium dimidiatum_hyalinum</i>
144	<i>Neoscytalidium</i> sp
145	<i>Ovatospora brasiliensis</i>
146	<i>Ovatospora</i> sp
147	<i>Paecilomyces lagunculariae</i>
148	<i>Paraphyton cookei</i>
149	<i>Paraphyton cookiellum</i>
150	<i>Penicillium aurantiogriseum</i>
151	<i>Penicillium brevicompactum</i>
152	<i>Penicillium camemberti_commune_cyclopium</i>



	Species/Group
153	<i>Penicillium canescens</i>
154	<i>Penicillium capsulatum</i>
155	<i>Penicillium chrysogenum</i>
156	<i>Penicillium citreonigrum</i>
157	<i>Penicillium citrinum</i>
158	<i>Penicillium corylophilum</i>
159	<i>Penicillium digitatum</i>
160	<i>Penicillium expansum</i>
161	<i>Penicillium fellutanum</i>
162	<i>Penicillium glabrum</i>
163	<i>Penicillium italicum</i>
164	<i>Penicillium menonorum</i>
165	<i>Penicillium nalgiovense</i>
166	<i>Penicillium namyslowskii</i>
167	<i>Penicillium olsonii</i>
168	<i>Penicillium onobense</i>
169	<i>Penicillium oxalicum</i>
170	<i>Penicillium pimiteouiense</i>
171	<i>Penicillium roqueforti</i>
172	<i>Penicillium singorense</i>
173	<i>Penicillium sp</i>
174	<i>Penicillium sp[2]</i>
175	<i>Penicillium turbatum</i>
176	<i>Penicillium verrucosum</i>
177	<i>Petriella setifera</i>
178	<i>Phaeoacremonium cinereum</i>
179	<i>Phialemoniopsis curvata</i>
180	<i>Phoma herbarum</i>
181	<i>Plectosphaerella cucumerina</i>
182	<i>Pseudogymnoascus pannorum</i>
183	<i>Purpureocillium lilacinum</i>
184	<i>Rasamsonia argillacea</i>
185	<i>Rhizomucor miehei</i>
186	<i>Rhizomucor pusillus</i>
187	<i>Rhizopus delemar</i>
188	<i>Rhizopus microsporus</i>
189	<i>Rhizopus oryzae</i>

	Species/Group
190	<i>Rhizopus stolonifer</i>
191	<i>Sarocladium kiliense</i>
192	<i>Sarocladium strictum</i>
193	<i>Scedosporium sp[5]</i>
194	<i>Schizophyllum commune</i>
195	<i>Scopulariopsis brevicaulis</i>
196	<i>Scytalidium sp</i>
197	<i>Sporothrix schenckii</i>
198	<i>Stachybotrys chartarum_chlorohalonata</i>
199	<i>Stachybotrys echinata</i>
200	<i>Syncephalastrum monosporum</i>
201	<i>Syncephalastrum racemosum</i>
202	<i>Talaromyces bacillisporus</i>
203	<i>Talaromyces diversus</i>
204	<i>Talaromyces duclauxii_wortmannii</i>
205	<i>Talaromyces funiculosus</i>
206	<i>Talaromyces islandicus</i>
207	<i>Talaromyces macrosporus</i>
208	<i>Talaromyces pseudostromaticus</i>
209	<i>Talaromyces ruber</i>
210	<i>Talaromyces rugulosus</i>
211	<i>Talaromyces sp</i>
212	<i>Talaromyces trachyspermus</i>
213	<i>Thanatephorus cucumeris</i>
214	<i>Trichoderma fertile</i>
215	<i>Trichoderma hamatum</i>
216	<i>Trichoderma harzianum</i>
217	<i>Trichoderma longibrachiatum_orientale</i>
218	<i>Trichoderma polysporum</i>
219	<i>Trichoderma reesei</i>
220	<i>Trichophyton mentagrophytes_group</i>
221	<i>Trichophyton rubrum_group</i>
222	<i>Trichophyton terrestre</i>
223	<i>Trichothecium roseum</i>
224	<i>Trichurus spiralis</i>
225	<i>Zopfiella karachiensis</i>



4 Renaming

Table 2: Renaming of reference entries

Filamentous Fungi Library Version 2022	Filamentous Fungi Library 2023	Justification
Curvularia sp[6] 025_34602_927 JMI	Curvularia sp[7] 025_34602_927 JMI	Since species in <i>Curvularia</i> sp[6] (<i>Curvularia clavata</i> , <i>Curvularia fallax</i> , <i>Curvularia hawaiiensis</i> , <i>Curvularia lunata</i> , <i>Curvularia pallescens</i> and <i>Curvularia verruculosa</i>) and the newly added species <i>Curvularia spicifera</i> cannot be separated by MALDI, they have been grouped together. Therefore, all mentioned species should be taken into consideration as a possible result.
Curvularia sp[6] 136_37245_1066 JMI	Curvularia sp[7] 136_37245_1066 JMI	
Curvularia sp[6] 181009_27 MUW	Curvularia sp[7] 181009_27 MUW	
Curvularia sp[6] 2012_23_A ALB	Curvularia sp[7] 2012_23_A ALB	
Curvularia sp[6] 2016_31_F IFS	Curvularia sp[7] 2016_31_F IFS	
Curvularia sp[6] 352_50243_2003 JMI	Curvularia sp[7] 352_50243_2003 JMI	
Curvularia sp[6] CBS 150_63 CBS	Curvularia sp[7] CBS 150_63 CBS	
Curvularia sp[6] CBS 156_35T CBS	Curvularia sp[7] CBS 156_35T CBS	
Curvularia sp[6] CBS 294_61 CBS	Curvularia sp[7] CBS 294_61 CBS	
Curvularia sp[6] CBS 408_96 CBS	Curvularia sp[7] CBS 408_96 CBS	
Curvularia sp[6] CBS 444_7 CBS_2	Curvularia sp[7] CBS 444_7 CBS_2	
Curvularia sp[6] CBS 539_70 CBS_2	Curvularia sp[7] CBS 539_70 CBS_2	
Curvularia sp[6] DSM 1157 DSM	Curvularia sp[7] DSM 1157 DSM	
Curvularia sp[6] DSM 62480 DSM	Curvularia sp[7] DSM 62480 DSM	
Curvularia sp[6] DSM 62482 DSM	Curvularia sp[7] DSM 62482 DSM	
Curvularia sp[6] F70 LLH	Curvularia sp[7] F70 LLH	
Penicillium camemberti DSM 1233 DSM	Penicillium camemberti_communis_cyclopium DSM 1233 DSM	Since species <i>Penicillium camemberti</i> and <i>Penicillium commune</i> and the



Filamentous Fungi Library Version 2022	Filamentous Fungi Library 2023	Justification
Penicillium camemberti DSM 1995 DSM	Penicillium camemberti_communis_cyclopium DSM 1995 DSM	newly added species <i>Penicillium cyclopium</i> cannot be separated by MALDI, they have been grouped together. Therefore, all mentioned species should be taken into consideration as a possible result.
Penicillium commune 1260 MPA	Penicillium camemberti_communis_cyclopium 1260 MPA	
Stachybotrys chartarum 17122224 LMU	Stachybotrys chartarum_chlorohalonata 17122224 LMU	Since species <i>Stachybotrys chartarum</i> and <i>Stachybotrys chlorohalonata</i> cannot be separated by MALDI, they have been grouped together. Therefore, all mentioned species should be taken into consideration as a possible result.
Stachybotrys chartarum 17122225 LMU	Stachybotrys chartarum_chlorohalonata 17122225 LMU	
Stachybotrys chartarum 192239 MPA	Stachybotrys chartarum_chlorohalonata 192239 MPA	
Stachybotrys chartarum 19523 MPA	Stachybotrys chartarum_chlorohalonata 19523 MPA	
Stachybotrys chartarum DSM 12880 DSM	Stachybotrys chartarum_chlorohalonata DSM 12880 DSM	
Stachybotrys chlorohalonata 18032500 LMU	Stachybotrys chartarum_chlorohalonata 18032500 LMU	
Stachybotrys chlorohalonata 19813 MPA	Stachybotrys chartarum_chlorohalonata 19813 MPA	
Talaromyces duclauxii CBS 187_89 CBS	Talaromyces duclauxii_wortmannii CBS 187_89 CBS	Since species <i>Talaromyces duclauxii</i> and <i>Talaromyces wortmannii</i> cannot be separated by MALDI. they have been grouped together. Therefore, all mentioned species should be taken into consideration as a possible result.
Talaromyces duclauxii CBS 323_48 CBS	Talaromyces duclauxii_wortmannii CBS 323_48 CBS	
Talaromyces duclauxii DSM 3826 DSM_2	Talaromyces duclauxii_wortmannii DSM 3826 DSM_2	
Talaromyces wortmannii 1271 MPA	Talaromyces duclauxii_wortmannii 1271 MPA	
Trichoderma longibrachiatum 17326100_1 MVD	Trichoderma longibrachiatum_orientale 17326100_1 MVD	Since species <i>Trichoderma longibrachiatum</i> and <i>Trichoderma orientale</i> cannot be separated by MALDI, they have been grouped together. Therefore, all mentioned species should be taken into consideration as a possible result.
Trichoderma longibrachiatum C18 SPF	Trichoderma longibrachiatum_orientale C18 SPF	
Trichoderma longibrachiatum DSM 63060 DSM	Trichoderma longibrachiatum_orientale DSM 63060 DSM	
Trichoderma orientale CBS 131488 CBS	Trichoderma longibrachiatum_orientale CBS 131488 CBS	
Trichoderma orientale DSM 16517 DSM	Trichoderma longibrachiatum_orientale DSM 16517 DSM	
Exophiala dermatitidis CBS 207_35 CBS	Exophiala dermatitidis CBS 207_35T CBS	Cosmetic change

5 Groups

The following table lists groups and their consolidated species within the MBT Filamentous Fungi Library 2023.

Table 3: Groups and their consolidated species

Group	Consolidated species
<i>Aspergillus flavus_oryzae_group</i>	<i>Aspergillus flavus</i> <i>Aspergillus oryzae</i>
<i>Aspergillus</i> sp[4] F45 LLH	<i>Aspergillus candidus</i> <i>Aspergillus campestris</i> <i>Aspergillus pragensis</i> <i>Aspergillus subalbidus</i>
<i>Curvularia</i> sp[7]	<i>Curvularia clavata</i> <i>Curvularia fallax</i> <i>Curvularia hawaiiensis</i> <i>Curvularia lunata</i> <i>Curvularia pallescens</i> <i>Curvularia spicifera</i> <i>Curvularia verruculosa</i>
<i>Fusarium cerealis_culmorum_group</i>	<i>Fusarium cerealis</i> <i>Fusarium culmorum</i>
<i>Penicillium camemberti_commune_cyclopium_group</i>	<i>Penicillium camemberti</i> <i>Penicillium commune</i> <i>Penicillium cyclopium</i>
<i>Penicillium</i> sp[2] 1331 MPA	<i>Penicillium aurantiocandidum</i> <i>Penicillium cellarum</i>
<i>Penicillium</i> sp[2] DSM 62843 DSM	<i>Penicillium hispanicum</i> <i>Penicillium implicatum</i>
<i>Scedosporium</i> sp[5]	<i>Scedosporium apiospermum</i> <i>Scedosporium aurantiacum</i> <i>Scedosporium boydii</i> <i>Scedosporium dehoogii</i> <i>Scedosporium minutisporum</i>
<i>Stachybotrys chartarum_chlorohalonata_group</i>	<i>Stachybotrys chartarum</i> <i>Stachybotrys chlorohalonata</i>
<i>Talaromyces duclauxii_wortmannii_group</i>	<i>Talaromyces duclauxii</i> <i>Talaromyces wortmannii</i>
<i>Trichoderma longibrachiatum_orientale_group</i>	<i>Trichoderma longibrachiatum</i> <i>Trichoderma orientale</i>
<i>Trichophyton mentagrophytes_group</i>	<i>Trichophyton benhamiae</i> <i>Trichophyton equinum</i> <i>Trichophyton erinaceid</i> <i>Trichophyton eriotrephon</i> <i>Trichophyton interdigitale</i> <i>Trichophyton mentagrophytes</i> <i>Trichophyton tonsurans</i> <i>Trichophyton verrucosum</i>



Group	Consolidated species
<i>Trichophyton rubrum_group</i>	<i>Trichophyton rubrum</i> <i>Trichophyton violaceum</i>



6 Changes to Matching Hints

Table 4: Matching Hint Changes

Species/Group	Action	MBT Filamentous Fungi Library 2023	Justification
<i>Aspergillus carneus</i>	Implement new Matching Hint	is a member of <i>Aspergillus</i> sect. <i>terri</i> group or closely related	Due to recent sequencing results the adaption of the Matching Hint is needed.
<i>Aspergillus ruber</i>	Implement new Matching Hint	is a member of <i>Aspergillus</i> sect. <i>aspergillus</i> group or closely related	Due to recent sequencing results the adaption of the Matching Hint is needed.
<i>Aspergillus terreus</i>	Implement new Matching Hint	is a member of <i>Aspergillus</i> sect. <i>terri</i> group or closely related	Due to recent sequencing results the adaption of the Matching Hint is needed.
<i>Cladosporium halotolerans</i>	Implement new Matching Hint	closely related to <i>Cladosporium sphaerospermum</i> and not definitely distinguishable at the moment	Due to recent sequencing results the adaption of the Matching Hint is needed.
<i>Cladosporium sphaerospermum</i>	Implement new Matching Hint	closely related to <i>Cladosporium halotolerans</i> and not definitely distinguishable at the moment	Due to recent sequencing results the adaption of the Matching Hint is needed.
<i>Fusarium cerealis</i>	Implement new Matching Hint	closely related to <i>Fusarium graminearum</i> , <i>Fusarium sambucinum</i> SC group and not definitely distinguishable at the moment	Due to recent sequencing results the adaption of the Matching Hint is needed.
<i>Fusarium culmorum</i>	Implement new Matching Hint	closely related to <i>Fusarium graminearum</i> , <i>Fusarium sambucinum</i> SC group and not definitely distinguishable at the moment	Due to recent sequencing results the adaption of the Matching Hint is needed.

7 Technical Information

7.1 Installation Instruction

- For **MBT Compass** the prerequisite is the use of the MBT Compass software (Build 4.1.100) and the MBT Compass Filamentous Fungi Module.
 - Execute the setup file: MaldiBiotyper_Library-Module_Filamentous-Fungi-V7.0.0.0_1021-MSPs_(RUO).exe
- Note:** Running setups requires administrator rights and can take several hours to complete. Please be aware that once the MSP library update was started, the progress must not be interrupted. Any interruption would leave the installed MSP library in an intermediate and therefore invalid state.
- After successful installation, the appropriate MBT client application will use the MBT Filamentous Fungi Library 2023 containing 1021 MSPs.
- Note:** Be aware that MSPs of the new library are assigned to the appropriate nodes in the taxonomy tree and are not available in the project tree. Accordingly, the Filamentous Fungi MSPs are accessed via the MSP library, if required.
- Note:** The installation log and a complete name list of the installed MSPs are available in the folder **C:\BDALSystemData\MBT_Setups**. Please provide all files that are available in the folder to the Bruker Support in case of problems or questions.
- For **MBT Compass HT** refer to the MBT HT Filamentous Fungi Module User Manual.

7.2 Compatibility of MALDI Biotyper software versions

MBT Filamentous Fungi Library 2023 can be used with the following MBT RUO software versions:

- MBT Compass
- MBT Compass HT



8 Appendix

8.1 Matching Hints

The following table lists the Matching Hints included in the MBT Filamentous Fungi Library 2023.

Table 5: Matching Hints

Species/Group	Matching Hint	References
<i>Alternaria alternata</i>	is a member of Alternaria sect. alternata group or closely related	Woudenberg, J.H.C., Groenewald, J.Z., Binder, M., Crous, P.W., 2013. Alternaria redefined. Stud. Mycol. 75, 171–212.
<i>Alternaria infectoria</i>	is a member of Alternaria sect. infectoriae group or closely related	Woudenberg, J.H.C., Groenewald, J.Z., Binder, M., Crous, P.W., 2013. Alternaria redefined. Studies in Mycology 75, 171–212.
<i>Arthrimum phaeospermum</i>	is a member of Arthrimum sect. apiospora group or closely related	Tian, X., Karunarathna, S.C., Mapook, A., Promputtha, I., Xu, J., Bao, D., Tibpromma, S., 2021. One New Species and Two New Host Records of Apiospora from Bamboo and Maize in Northern Thailand with Thirteen New Combinations. Life 11, 1071.
<i>Arthroderma borellii</i>	synonym of Arthroderma amazonicum	N/A
<i>Arthroderma gertleri</i>	closely related to Arthroderma gloriae and not definitely distinguishable at the moment	N/A
<i>Arthroderma gloriae</i>	closely related to Arthroderma gertleri and not definitely distinguishable at the moment	N/A



Species/Group	Matching Hint	References
<i>Aspergillus brasiliensis</i>	is a member of <i>Aspergillus</i> sect. <i>nigri</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus carneus</i>	is a member of <i>Aspergillus</i> sect. <i>terri</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus calidoustus</i>	is a member of <i>Aspergillus</i> sect. <i>usti</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus flavus_oryzae_group</i>	is a member of <i>Aspergillus</i> sect. <i>flavi</i> group or closely related	Visagie, C.M., Houbraken, J., 2020. Updating the taxonomy of <i>Aspergillus</i> in South Africa. <i>Stud. Mycol.</i> 95, 253–292.
<i>Aspergillus fumigatus</i>	synonym of <i>Aspergillus neoellipticus</i> group	N/A



Species/Group	Matching Hint	References
<i>Aspergillus japonicus</i>	is a member of <i>Aspergillus</i> sect. <i>nigri</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus lentulus</i>	is a member of <i>Aspergillus</i> sect. <i>fumigati</i> group or closely related	Visagie, C.M., Houbraken, J., 2020. Updating the taxonomy of <i>Aspergillus</i> in South Africa. <i>Stud. Mycol.</i> 95, 253–292.
<i>Aspergillus montevidensis</i>	synonym of <i>Aspergillus amstelodami</i>	N/A
<i>Aspergillus niger</i>	is a member of <i>Aspergillus</i> sect. <i>nigri</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus ochraceus</i>	is a member of <i>Aspergillus</i> sect. <i>circumdati</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus parasiticus</i>	is a member of <i>Aspergillus</i> sect. <i>flavi</i> group or closely related	Visagie, C.M., Houbraken, J., 2020. Updating the taxonomy of <i>Aspergillus</i> in South Africa. <i>Stud. Mycol.</i> 95, 253–292.



Species/Group	Matching Hint	References
<i>Aspergillus ruber</i>	is a member of <i>Aspergillus</i> sect. <i>aspergillus</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus sclerotiorum</i>	is a member of <i>Aspergillus</i> sect. <i>circumdati</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus sydowii</i>	is a member of <i>Aspergillus</i> sect. <i>nidulantes</i> group or closely related	Visagie, C.M., Houbraken, J., 2020. Updating the taxonomy of <i>Aspergillus</i> in South Africa. <i>Stud. Mycol.</i> 95, 253–292.
<i>Aspergillus tamarii</i>	is a member of <i>Aspergillus</i> sect. <i>flavi</i> group or closely related	Visagie, C.M., Houbraken, J., 2020. Updating the taxonomy of <i>Aspergillus</i> in South Africa. <i>Stud. Mycol.</i> 95, 253–292.
<i>Aspergillus terreus</i>	is a member of <i>Aspergillus</i> sect. <i>terri</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.- C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus tritici</i>	is a member of <i>Aspergillus</i> sect. <i>candidi</i> group or closely related	Varga, J., Frisvad, J.C., Samson, R.A., 2007. Polyphasic taxonomy of <i>Aspergillus</i> section <i>Candidi</i> based on molecular, morphological and physiological data. <i>Studies in Mycology</i> 59, 75–88.



Species/Group	Matching Hint	References
<i>Aspergillus unguis</i>	is a member of <i>Aspergillus</i> sect. <i>nidulantes</i> group or closely related	Visagie, C.M., Houbraken, J., 2020. Updating the taxonomy of <i>Aspergillus</i> in South Africa. <i>Stud. Mycol.</i> 95, 253–292.
<i>Aspergillus ustus</i>	is a member of <i>Aspergillus</i> sect. <i>usti</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.-C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Aspergillus versicolor</i>	is a member of <i>Aspergillus</i> sect. <i>nidulantes</i> group or closely related	Visagie, C.M., Houbraken, J., 2020. Updating the taxonomy of <i>Aspergillus</i> in South Africa. <i>Stud. Mycol.</i> 95, 253–292.
<i>Aspergillus westerdijkiae</i>	is a member of <i>Aspergillus</i> sect. <i>circumdati</i> group or closely related	Houbraken, J., Kocsubé, S., Visagie, C.M., Yilmaz, N., Wang, X.-C., Meijer, M., Kraak, B., Hubka, V., Bensch, K., Samson, R.A., Frisvad, J.C., 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. <i>Studies in Mycology</i> 95, 5–169.
<i>Boeremia exigua</i>	is a member of <i>Boeremia</i> group or closely related	Marin-Felix, Y., Groenewald, J.Z., Cai, L., Chen, Q., Marincowitz, S., Barnes, I., Bensch, K., Braun, U., Camporesi, E., Damm, U., de Beer, Z.W., Dissanayake, A., Edwards, J., Giraldo, A., Hernández-Restrepo, M., Hyde, K.D., Jayawardena, R.S., Lombard, L., Luangsa-ard, J., McTaggart, A.R., Rossman, A.Y., Sandoval-Denis, M., Shen, M., Shivas, R.G., Tan, Y.P., van der Linde, E.J., Wingfield, M.J., Wood, A.R., Zhang, J.Q., Zhang, Y., Crous, P.W., 2017. Genera of phytopathogenic fungi: GOPHY 1. <i>Studies in Mycology</i> 86, 99–216.



Species/Group	Matching Hint	References
<i>Botrytis aclada</i>	is a member of Botrytis 1 group or closely related	Walker, A.-S., 2016. Diversity Within and Between Species of Botrytis, in: Fillinger, S., Elad, Y. (Eds.), Botrytis – the Fungus, the Pathogen and Its Management in Agricultural Systems. Springer International Publishing, Cham, pp. 91–125.
<i>Botrytis cinerea</i>	is a member of Botrytis 1 group or closely related	Walker, A.-S., 2016. Diversity Within and Between Species of Botrytis, in: Fillinger, S., Elad, Y. (Eds.), Botrytis – the Fungus, the Pathogen and Its Management in Agricultural Systems. Springer International Publishing, Cham, pp. 91–125.
<i>Byssoschlamys fulva</i>	is a member of Byssoschlamys 1 group or closely related	Samson, R.A., Houbraken, J., Varga, J., Frisvad, J.C., 2009. Polyphasic taxonomy of the heat resistant ascomycete genus Byssoschlamys and its Paecilomyces anamorphs. Pers - Int Mycol J 22, 14–27.
<i>Byssoschlamys nivea</i>	is a member of Byssoschlamys 1 group or closely related	Samson, R.A., Houbraken, J., Varga, J., Frisvad, J.C., 2009. Polyphasic taxonomy of the heat resistant ascomycete genus Byssoschlamys and its Paecilomyces anamorphs. Pers - Int Mycol J 22, 14–27.
<i>Byssoschlamys spectabilis</i>	synonym of Paecilomyces variotii	Houbraken, Jos, János Varga, Emilia Rico-Munoz, Shawn Johnson, and Robert A. Samson. 2008. Sexual Reproduction as the Cause of Heat Resistance in the Food Spoilage Fungus Byssoschlamys Spectabilis (Anamorph Paecilomyces Variotii). Applied and Environmental Microbiology 74(5):1613–19. doi: 10.1128/AEM.01761-07.
<i>Chaetomium globosum</i>	is a member of Chaetomium 2 group or closely related	Wang, X.W., Lombard, L., Groenewald, J.Z., Li, J., Videira, S.I.R., Samson, R.A., Liu, X.Z., Crous, P.W., 2016. Phylogenetic reassessment of the Chaetomium globosum species complex. Pers - Int Mycol J 36, 83–133.



Species/Group	Matching Hint	References
<i>Cladosporium cladosporioides</i>	is a member of <i>Cladosporium cladosporioides</i> complex	Sandoval-Denis, M., Gené, J., Sutton, D.A., Wiederhold, N.P., Cano-Lira, J.F., Guarro, J., 2016. New species of <i>Cladosporium</i> associated with human and animal infections. Pers - Int Mycol J 36, 281–298.
<i>Cladosporium halotolerans</i>	closely related to <i>Cladosporium sphaerospermum</i> and not definitely distinguishable at the moment	N/A
<i>Cladosporium herbarum</i>	is a member of <i>Cladosporium herbarum</i> complex	Sandoval-Denis, M., Gené, J., Sutton, D.A., Wiederhold, N.P., Cano-Lira, J.F., Guarro, J., 2016. New species of <i>Cladosporium</i> associated with human and animal infections. Pers - Int Mycol J 36, 281–298.
<i>Cladosporium sphaerospermum</i>	closely related to <i>Cladosporium halotolerans</i> and not definitely distinguishable at the moment	N/A
<i>Cladosporium macrocarpum</i>	is a member of <i>Cladosporium herbarum</i> complex	Sandoval-Denis, M., Gené, J., Sutton, D.A., Wiederhold, N.P., Cano-Lira, J.F., Guarro, J., 2016. New species of <i>Cladosporium</i> associated with human and animal infections. Pers - Int Mycol J 36, 281–298.
<i>Clonostachys rosea</i>	is a member of <i>Clonostachys</i> sect. <i>clonostachys</i> group or closely related	Forin, N., Vizzini, A., Nigris, S., Ercole, E., Voyron, S., Girlanda, M., Baldan, B., 2020. Illuminating type collections of nectriaceous fungi in Saccardo's fungarium. <i>persoonia</i> 45, 221–249.
<i>Colletotrichum gloeosporioides</i>	synonym of <i>Colletotrichum aeschynomenes</i>	N/A
<i>Coniochaeta luteorubra</i>	synonym of <i>Coniochaeta hoffmannii</i>	N/A
<i>Dichotomopilus funicola</i>	is a member of <i>Dichotomopilus</i> gen. nov. group or closely related	Wang, X.W., Houbraken, J., Groenewald, J.Z., Meijer, M., Andersen, B., Nielsen, K.F., Crous, P.W., Samson, R.A., 2016. Diversity and taxonomy of <i>Chaetomium</i> and <i>chaetomium</i> -like fungi from indoor environments. <i>Studies in Mycology</i> 84, 145– 224.



Species/Group	Matching Hint	References
<i>Didymella glomerata</i>	is a member of <i>Didymella peyronellaea</i> - clade group or closely related	Scarpari, M., Vitale, S., Di Giambattista, G., Luongo, L., De Gregorio, T., Schreiber, G., Petrucci, M., Belisario, A., Voglmayr, H., 2020. <i>Didymella corylicola</i> sp. nov., a new fungus associated with hazelnut fruit development in Italy. <i>Mycol Progress</i> 19, 317– 328.
<i>Didymella pomorum</i>	is a member of <i>Didymella</i> sect. <i>didymella</i> group or closely related	Keirnan, E.C., Tan, Y.P., Laurence, M.H., Mertin, A.A., Liew, E.C.Y., Summerell, B.A., Shivas, R.G., 2021. Cryptic diversity found in <i>Didymellaceae</i> from Australian native legumes. <i>MC</i> 78, 1–20.
<i>Epicoccum nigrum</i>	is a member of <i>Epicoccum</i> group or closely related	Chen, Q., Hou, L.W., Duan, W.J., Crous, P.W., Cai, L., 2017. <i>Didymellaceae</i> revisited. <i>Studies in Mycology</i> 87, 105–159.
<i>Fusarium cerealis</i>	closely related to <i>Fusarium graminearum</i> , <i>Fusarium sambucinum</i> SC group and not definitely distinguishable at the moment	Stępniewska, H., Jankowiak, R., Bilański, P., Hausner, G., 2021. Structure and Abundance of <i>Fusarium</i> Communities Inhabiting the Litter of Beech Forests in Central Europe. <i>Forests</i> 12, 811.
<i>Fusarium culmorum</i>	closely related to <i>Fusarium graminearum</i> , <i>Fusarium sambucinum</i> SC group and not definitely distinguishable at the moment	Stępniewska, H., Jankowiak, R., Bilański, P., Hausner, G., 2021. Structure and Abundance of <i>Fusarium</i> Communities Inhabiting the Litter of Beech Forests in Central Europe. <i>Forests</i> 12, 811.
<i>Fusarium delphinoides</i>	is a member of <i>Fusarium dimerum</i> species group or closely related	Schroers, H.-J., O'Donnell, K., Lamprecht, S.C., Kammeyer, P.L., Johnson, S., Sutton, D.A., Rinaldi, M.G., Geiser, D.M., Summerbell, R.C., 2009. Taxonomy and phylogeny of the <i>Fusarium dimerum</i> species group. <i>Mycologia</i> 101, 44–70.
<i>Fusarium dimerum</i>	is a member of <i>Fusarium dimerum</i> species group or closely related	Schroers, H.-J., O'Donnell, K., Lamprecht, S.C., Kammeyer, P.L., Johnson, S., Sutton, D.A., Rinaldi, M.G., Geiser, D.M., Summerbell, R.C., 2009. Taxonomy and phylogeny of the <i>Fusarium dimerum</i> species group. <i>Mycologia</i> 101, 44–70.



Species/Group	Matching Hint	References
<i>Fusarium graminearum</i>	closely related to <i>Fusarium cerealis_culmorum</i> , <i>Fusarium sambucinum</i> SC group and not definitely distinguishable at the moment	Stępniewska, H., Jankowiak, R., Bilański, P., Hausner, G., 2021. Structure and Abundance of <i>Fusarium</i> Communities Inhabiting the Litter of Beech Forests in Central Europe. <i>Forests</i> 12, 811.
<i>Fusarium incarnatum</i>	is a member of <i>Fusarium fujikuroi</i> species complex	Villani, A., Proctor, R.H., Kim, H.-S., Brown, D.W., Logrieco, A.F., Amatulli, M.T., Moretti, A., Susca, A., 2019. Variation in secondary metabolite production potential in the <i>Fusarium incarnatum-equiseti</i> species complex revealed by comparative analysis of 13 genomes. <i>BMC Genomics</i> 20, 314.
<i>Fusarium oxysporum</i>	closely related to <i>Fusarium proliferatum</i> , <i>Fusarium verticillioides</i> and not definitely distinguishable at the moment	N/A
<i>Fusarium petroliphilum</i>	synonym of <i>Fusarium solani</i> var. <i>petroliphilum</i>	N/A
<i>Fusarium proliferatum</i>	is a member of <i>Fusarium fujikuroi</i> species complex	Villani, A., Proctor, R.H., Kim, H.-S., Brown, D.W., Logrieco, A.F., Amatulli, M.T., Moretti, A., Susca, A., 2019. Variation in secondary metabolite production potential in the <i>Fusarium incarnatum-equiseti</i> species complex revealed by comparative analysis of 13 genomes. <i>BMC Genomics</i> 20, 314.
<i>Fusarium solani</i>	is a member of <i>Fusarium solani</i> complex	Al-Hatmi, Abdullah M. S., Sarah A. Ahmed, Anne D. van Diepeningen, Miranda Drogari-Apiranthitou, Paul E. Verweij, Jacques F. Meis, and G. Sybren de Hoog. 2018. <i>Fusarium Metavorans</i> Sp. Nov.: The Frequent Opportunist 'FSSC6.' <i>Medical Mycology</i> 56:S144–52. doi: 10.1093/mmy/myx107.



Species/Group	Matching Hint	References
<i>Fusarium sporotrichioides</i>	is a member of <i>Fusarium sambucinum</i> group or closely related	Villani, A., Proctor, R.H., Kim, H.-S., Brown, D.W., Logrieco, A.F., Amatulli, M.T., Moretti, A., Susca, A., 2019. Variation in secondary metabolite production potential in the <i>Fusarium incarnatum-equiseti</i> species complex revealed by comparative analysis of 13 genomes. BMC Genomics 20, 314.
<i>Fusarium verticillioides</i>	is a member of <i>Fusarium fujikuroi</i> species complex	Villani, A., Proctor, R.H., Kim, H.-S., Brown, D.W., Logrieco, A.F., Amatulli, M.T., Moretti, A., Susca, A., 2019. Variation in secondary metabolite production potential in the <i>Fusarium incarnatum-equiseti</i> species complex revealed by comparative analysis of 13 genomes. BMC Genomics 20, 314.
<i>Lomentospora prolificans</i>	closely related to <i>Petriella setifera</i> and not definitely distinguishable at the moment	N/A
<i>Monascus ruber</i>	is a member of <i>Monascus</i> section <i>rubri</i> group or closely related	Barbosa, R. N., S. L. Leong, O. Vinnere-Pettersson, A. J. Chen, C. M. Souza-Motta, J. C. Frisvad, R. A. Samson, N. T. Oliveira, and J. Houbraken. 2017. Phylogenetic Analysis of <i>Monascus</i> and New Species from Honey, Pollen and Nests of Stingless Bees. Studies in Mycology 86(1):29-51. doi: 10.1016/j.simyco.2017.04.001.
<i>Mortierella angusta</i>	is a member of <i>Mortierella angusta</i> group or closely related	Wagner, L., Stielow, B., Hoffmann, K., Petkovits, T., Papp, T., Vágvolgyi, C., de Hoog, G.S., Verkley, G., Voigt, K., 2013. A comprehensive molecular phylogeny of the Mortierellales (Mortierellomycotina) based on nuclear ribosomal DNA. Pers - Int Mycol J 30, 77–93.
<i>Neoscytalidium</i> sp	synonym of <i>Scytalidium</i> sp.	N/A



Species/Group	Matching Hint	References
<i>Ovatospora brasiliensis</i>	is a member of <i>Ovatospora</i> gen. nov. group or closely related	Wang, X.W., Houbraken, J., Groenewald, J.Z., Meijer, M., Andersen, B., Nielsen, K.F., Crous, P.W., Samson, R.A., 2016. Diversity and taxonomy of <i>Chaetomium</i> and chaetomium-like fungi from indoor environments. <i>Stud. Mycol.</i> 84, 145–224.
<i>Ovatospora</i> sp	synonym of <i>Chaetomium</i> sp.	N/A
<i>Paecilomyces lagunculariae</i>	synonym of <i>Byssochlamys nivea</i>	N/A
<i>Penicillium aurantiogriseum</i>	is a member of <i>Penicillium</i> clade 15-23 group or closely related	Houbraken, J., Samson, R.A., 2011. Phylogeny of <i>Penicillium</i> and the segregation of <i>Trichocomaceae</i> into three families. <i>Studies in Mycology</i> 70, 1–51.
<i>Penicillium brevicompactum</i>	is a member of <i>Penicillium</i> sect. <i>brevicompacta</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium camemberti</i>	is a member of <i>Penicillium</i> sect. <i>fasciculata</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.



Species/Group	Matching Hint	References
<i>Penicillium chrysogenum</i>	is a member of <i>Penicillium</i> sect. <i>chrysogena</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium citreonigrum</i>	is a member of <i>Penicillium</i> sect. <i>exilicaulis</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium citrinum</i>	is a member of <i>Penicillium</i> sect. <i>citrina</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium commune</i>	is a member of <i>Penicillium</i> sect. <i>fasciculata</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.



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<i>Penicillium corylophilum</i>	is a member of <i>Penicillium</i> sect. <i>exilicaulis</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium cyclopium</i>	is a member of <i>Penicillium</i> sect. <i>fasciculata</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium digitatum</i>	is a member of <i>Penicillium</i> clade 15-23 group or closely related	Houbraken, J., Samson, R.A., 2011. Phylogeny of <i>Penicillium</i> and the segregation of <i>Trichocomaceae</i> into three families. <i>Studies in Mycology</i> 70, 1–51.
<i>Penicillium expansum</i>	is a member of <i>Penicillium</i> clade 15-23 group or closely related	Houbraken, J., Samson, R.A., 2011. Phylogeny of <i>Penicillium</i> and the segregation of <i>Trichocomaceae</i> into three families. <i>Studies in Mycology</i> 70, 1–51.
<i>Penicillium fellutanum</i>	is a member of <i>Penicillium</i> sect. <i>charlesii</i> / sect. <i>sclerotiora</i> group or closely related	Houbraken, J., Samson, R.A., 2011. Phylogeny of <i>Penicillium</i> and the segregation of <i>Trichocomaceae</i> into three families. <i>Studies in Mycology</i> 70, 1–51.
<i>Penicillium glabrum</i>	is a member of <i>Penicillium glabrum</i> complex	Barreto, M. C., Houbraken, J., Samson, R.A., Frisvad, J.C., San- Romão, M.V., and Barreto, M.C., Houbraken, J., Samson, R.A., Frisvad, J.C., San- Romão, M.V. 2011. Taxonomic Studies of the <i>Penicillium glabrum</i> Complex and the Description of a New Species <i>P. subericola</i> . <i>Fungal Diversity</i> 49:23–33. doi: 10.1007/s13225-011-0090-4.



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<i>Penicillium italicum</i>	is a member of <i>Penicillium</i> clade 15-23 group or closely related	Houbraken, J., Samson, R.A., 2011. Phylogeny of <i>Penicillium</i> and the segregation of Trichocomaceae into three families. <i>Studies in Mycology</i> 70, 1–51.
<i>Penicillium menonorum</i>	closely related to <i>Penicillium</i> sect. <i>exilicaulis</i> and not definitely distinguishable at the moment	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium namyslowskii</i>	is a member of <i>Penicillium</i> sect. <i>exilicaulis</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium olsonii</i>	is a member of <i>Penicillium</i> sect. <i>brevicompacta</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium onobense</i>	is a member of <i>Penicillium</i> ser. <i>simplicissima</i> group or closely related	Hotta, Y., Sato, J., Sato, H., Hosoda, A., Tamura, H., 2011. Classification of the Genus <i>Bacillus</i> Based on MALDI-TOF MS Analysis of Ribosomal Proteins Coded in S10 and <i>spc</i> Operons. <i>J. Agric. Food Chem.</i> 110415093730042.



Species/Group	Matching Hint	References
<i>Penicillium pimateouiense</i>	is a member of <i>Penicillium</i> sect. <i>exilicaulis</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium roqueforti</i>	is a member of <i>Penicillium</i> sect. <i>roquefortorum</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium singorense</i>	is a member of <i>Penicillium</i> ser. <i>dalearum</i> group or closely related	Hotta, Y., Sato, J., Sato, H., Hosoda, A., Tamura, H., 2011. Classification of the Genus <i>Bacillus</i> Based on MALDI-TOF MS Analysis of Ribosomal Proteins Coded in S10 and <i>spc</i> Operons. <i>J. Agric. Food Chem.</i> 110415093730042.
<i>Penicillium turbatum</i>	is a member of <i>Penicillium</i> sect. <i>turbata</i> group or closely related	Houbraken, J., S. Kocsubé, C. M. Visagie, N. Yilmaz, X. C. Wang, M. Meijer, B. Kraak, V. Hubka, K. Bensch, R. A. Samson, and J. C. Frisvad. 2020. Classification of <i>Aspergillus</i> , <i>Penicillium</i> , <i>Talaromyces</i> and Related Genera (Eurotiales): An Overview of Families, Genera, Subgenera, Sections, Series and Species. <i>Studies in Mycology</i> 95:5–169. doi: 10.1016/j.simyco.2020.05.002.
<i>Penicillium verrucosum</i>	is a member of <i>Penicillium</i> clade 15-23 group or closely related	Houbraken, J., Samson, R.A., 2011. Phylogeny of <i>Penicillium</i> and the segregation of <i>Trichocomaceae</i> into three families. <i>Studies in Mycology</i> 70, 1–51.



Species/Group	Matching Hint	References
<i>Petriella setifera</i>	closely related to Lomentospora prolificans and not definitely distinguishable at the moment	Vebliza, Y., W. Sjamsuridzal, A. Oetari, I. Santoso, and I. G. Roosheroe. 2018. Re-Identification of Five Strains of Rhizopus Arrhizus from Tempeh Based on ITS Regions of RDNA Sequence Data. P. 020167 in. Bali, Indonesia.
<i>Phialemoniopsis curvata</i>	synonym of Thyridium curvatum	Sugita, Ryosuke, and Kazuaki Tanaka. 2022. Thyridium Revised: Synonymisation of Phialemoniopsis under Thyridium and Establishment of a New Order, Thyridiales. MycoKeys 86:147–76. doi: 10.3897/mycokeys.86.78989.
<i>Plectosphaerella cucumerina</i>	is a member of Plectosphaerella 1 group or closely related	Su, Lei, Hui Deng, and Yong-Chun Niu. 2017. Phylogenetic Analysis of Plectosphaerella Species Based on Multi-Locus DNA Sequences and Description of P. Sinensis Sp. Nov. Mycol Progress 16:823–29. doi: 10.1007/s11557-017-1319-8.
<i>Rasamsonia argillacea</i>	is a member of Rasamsonia clade a group or closely related	Houbraken, J., Giraud, S., Meijer, M., Bertout, S., Frisvad, J.C., Meis, J.F., Bouchara, J.P., Samson, R.A., 2013. Taxonomy and Antifungal Susceptibility of Clinically Important Rasamsonia Species. J Clin Microbiol 51, 22–30.
<i>Rhizopus oryzae</i>	synonym of Rhizopus arrhizus	Vebliza, Y., W. Sjamsuridzal, A. Oetari, I. Santoso, and I. G. Roosheroe. 2018. Re-Identification of Five Strains of Rhizopus Arrhizus from Tempeh Based on ITS Regions of RDNA Sequence Data. P. 020167 in. Bali, Indonesia.
<i>Scopulariopsis brevicaulis</i>	is a member of Scopulariopsis sec. scopulariopsis group or closely related	Sandoval-Denis, M., Gené, J., Sutton, D.A., Cano-Lira, J.F., de Hoog, G.S., Decock, C.A., Wiederhold, N.P., Guarro, J., 2016. Redefining Microascus, Scopulariopsis and allied genera. Pers - Int Mycol J 36, 1–36.



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<i>Sporothrix schenckii</i>	is a member of <i>Sporothrix</i> species strains group or closely related	Zhang, Y., Hagen, F., Stielow, B., Rodrigues, A.M., Samerpitak, K., Zhou, X., Feng, P., Yang, L., Chen, M., Deng, S., Li, S., Liao, W., Li, R., Li, F., Meis, J.F., Guarro, J., Teixeira, M., Al-Zahrani, H.S., de Camargo, Z.P., Zhang, L., de Hoog, G.S., 2015. Phylogeography and evolutionary patterns in <i>Sporothrix</i> spanning more than 14 000 human and animal case reports. <i>Pers - Int Mycol J</i> 35, 1–20.
<i>Stachybotrys chartarum</i>	closely related to <i>Stachybotrys chlorohalonata</i> and not definitely distinguishable at the moment	N/A
<i>Stachybotrys chlorohalonata</i>	closely related to <i>Stachybotrys chartarum</i> and not definitely distinguishable at the moment	N/A
<i>Stachybotrys echinata</i>	is a member of <i>Stachybotryaceae</i> group or closely related	Tibpromma, S., Boonmee, S., Wijayawardene, N.N., Maharachchikumbura, S.S.N., McKenzie, E.H.C., Bahkali, A.H., E.B. Gareth, J.E.B., Hyde, K.D., Promputtha, I., 2016. The holomorph of <i>arasarcopodium</i> (<i>Stachybotryaceae</i>), introducing <i>P. pandanicola</i> sp. nov. on <i>Pandanus</i> sp. <i>Phytotaxa</i> 266, 250.
<i>Talaromyces duclauxii</i>	is a member of <i>Talaromyces</i> sect. <i>talaromyces</i> group or closely related	Sun, B.-D., Chen, A.J., Houbraken, J., Frisvad, J.C., Wu, W.-P., Wei, H.-L., Zhou, Y.-G., Jiang, X.-Z., Samson, R.A., 2020. New section and species in <i>Talaromyces</i> . <i>MC</i> 68, 75–113.
<i>Talaromyces funiculosus</i>	is a member of <i>Talaromyces</i> sect. <i>talaromyces</i> group or closely related	Sun, B.-D., Chen, A.J., Houbraken, J., Frisvad, J.C., Wu, W.-P., Wei, H.-L., Zhou, Y.-G., Jiang, X.-Z., Samson, R.A., 2020. New section and species in <i>Talaromyces</i> . <i>MC</i> 68, 75–113.
<i>Talaromyces macrosporus</i>	is a member of <i>Talaromyces</i> sect. <i>talaromyces</i> group or closely related	Sun, B.-D., Chen, A.J., Houbraken, J., Frisvad, J.C., Wu, W.-P., Wei, H.-L., Zhou, Y.-G., Jiang, X.-Z., Samson, R.A., 2020. New section and species in <i>Talaromyces</i> . <i>MC</i> 68, 75–113.



Species/Group	Matching Hint	References
<i>Talaromyces pseudostromaticus</i>	is a member of Talaromyces sect. purpurei group or closely related	Sun, B.-D., Chen, A.J., Houbraken, J., Frisvad, J.C., Wu, W.-P., Wei, H.-L., Zhou, Y.-G., Jiang, X.-Z., Samson, R.A., 2020. New section and species in Talaromyces. MC 68, 75–113.
<i>Talaromyces ruber</i>	is a member of Talaromyces sect. talaromyces group or closely related	Sun, B.-D., Chen, A.J., Houbraken, J., Frisvad, J.C., Wu, W.-P., Wei, H.-L., Zhou, Y.-G., Jiang, X.-Z., Samson, R.A., 2020. New section and species in Talaromyces. MC 68, 75–113.
<i>Talaromyces rugulosus</i>	is a member of Talaromyces sect. islandici group or closely related	Sun, B.-D., Chen, A.J., Houbraken, J., Frisvad, J.C., Wu, W.-P., Wei, H.-L., Zhou, Y.-G., Jiang, X.-Z., Samson, R.A., 2020. New section and species in Talaromyces. MC 68, 75–113.
<i>Talaromyces trachyspermus</i>	is a member of Talaromyces sect. trachyspermi group or closely related	Sun, B.-D., Chen, A.J., Houbraken, J., Frisvad, J.C., Wu, W.-P., Wei, H.-L., Zhou, Y.-G., Jiang, X.-Z., Samson, R.A., 2020. New section and species in Talaromyces. MycoKeys 68, 75–113.
<i>Talaromyces wortmannii</i>	is a member of Talaromyces sect. talaromyces group or closely related	Sun, B.-D., Chen, A.J., Houbraken, J., Frisvad, J.C., Wu, W.-P., Wei, H.-L., Zhou, Y.-G., Jiang, X.-Z., Samson, R.A., 2020. New section and species in Talaromyces. MycoKeys 68, 75–113.
<i>Thanatephorus cucumeris</i>	synonym of Bjerkandera adusta	N/A
<i>Trichoderma fertile</i>	is a member of Trichoderma sect. semiorbis group or closely related	Jaklitsch, W.M., Voglmayr, H., 2015. Biodiversity of Trichoderma (Hypocreaceae) in Southern Europe and Macaronesia. Stud. Mycol. 80, 1–87.
<i>Trichoderma hamatum</i>	is a member of Trichoderma section hamatum* clade group or closely related	Druzhinina, I.S., Kopchinskiy, A.G., Kubicek, C.P., 2006. The first 100 Trichoderma species characterized by molecular data. Mycoscience 47, 55–64.



Species/Group	Matching Hint	References
<i>Trichoderma harzianum</i>	is a member of Trichoderma harzianum species complex	Chaverri, P., Branco-Rocha, F., Jaklitsch, W., Gazis, R., Degenkolb, T., Samuels, G.J., 2015. Systematics of the Trichoderma harzianum species complex and the re-identification of commercial biocontrol strains. Mycologia 107, 558–590.
<i>Trichoderma longibrachiatum</i>	is a member of Trichoderma section longibrachiatum group or closely related	Druzhinina, I.S., Kopchinskiy, A.G., Kubicek, C.P., 2006. The first 100 Trichoderma species characterized by molecular data. Mycoscience 47, 55–64.
<i>Trichoderma orientale</i>	is a member of Trichoderma section longibrachiatum group or closely related	Druzhinina, I.S., Kopchinskiy, A.G., Kubicek, C.P., 2006. The first 100 Trichoderma species characterized by molecular data. Mycoscience 47, 55–64.
<i>Trichoderma polysporum</i>	is a member of Trichoderma section polysporum clade group or closely related	Jaklitsch, W.M., Voglmayr, H., 2015. Biodiversity of Trichoderma (Hypocreaceae) in Southern Europe and Macaronesia. Stud. Mycol. 80, 1–87.
<i>Trichoderma reesei</i>	is a member of Trichoderma section longibrachiatum group or closely related	Jaklitsch, W.M., Voglmayr, H., 2015. Biodiversity of Trichoderma (Hypocreaceae) in Southern Europe and Macaronesia. Stud. Mycol. 80, 1–87.
<i>Trichophyton mentagrophytes_group</i>	is a member of Trichophyton mentagrophytes group or closely related	Gräser, Y., Kuijpers, A.F.A., Presber, W., De Hoog, G.S., 2008. Molecular taxonomy of Trichophyton mentagrophytes and T. tonsurans: Taxonomy of dermatophytes. Med Mycol 37, 315–330.
<i>Trichophyton rubrum_group</i>	is a member of Trichophyton rubrum group or closely related	Gräser, Y., Kuijpers, A.F.A., Presber, W., de Hoog, G.S., 2000. Molecular Taxonomy of the Trichophyton rubrum Complex. Journal of Clinical Microbiology 38, 3329–3336.
<i>Trichophyton terrestre</i>	synonym of Arthroderma quadrifidum	N/A



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<i>Trichurus spiralis</i>	is a member of Cephalotrichum gorgonifer group or closely related	Sandoval-Denis, M., Guarro, J., Cano-Lira, J.F., Sutton, D.A., Wiederhold, N.P., de Hoog, G.S., Abbott, S.P., Decock, C., Sigler, L., Gené, J., 2016. Phylogeny and taxonomic revision of Microascaceae with emphasis on synnematosus fungi. Studies in Mycology 83, 193–233.



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